

Course Unit	Virology and Animal Cell Culture			Field of study	Veterinary Technology		
Bachelor in	Veterinary Nursing			School	School of Agriculture		
Academic Year	2019/2020	Year of study	2	Level	1-2	ECTS credits	6.0
Туре	Semestral	Semester	2	Code	9085-408-2205-00-19		
Workload (hours)	162	Contact hours	T 30 TP	- PL 30 T	c - s -	E - OT	20 0 -
			T - Lectures; TP - Lectures a	and problem-solving; PL - Problem-	solving, project or laboratory; TC	- Fieldwork; S - Seminar; E - Place	ment; OT - Tutorial; O - Other
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Name(s) of lecturer(s) Hélder Miranda Pires Quintas, Joaquina Teresa Gaudêncio Dias, Manuel Ricardo Costa Calhelha

### Learning outcomes and competences

- At the end of the course unit the learner is expected to be able to:

  1. Learn the fundaments of viral taxonomy, viral replication, genetics and evolution, virus cell-interactions, mechanism of infection and viral spread, determinants of viral virulence and host resistance

- 2. Learn the fundaments of immune response to viral infections and pathogenesis of viral disease.

  3. Be familiar with laboratory diagnosis of viral diseases.

  4. Understand the concepts of surveillance, prevention, control and eradication of viral diseases with special attention to zoonotic agents.

  5. Learn the fundaments of prions: agents of transmissible spongiform encephalopathies.

  6. Know the most important applications for animal cell culture.

  7. Perform correctly the main procedures used in animal cell culture.

# Prerequisites

Before the course unit the learner is expected to be able to:

The students should have previous knowledge of biochemistry, physiology, histology and genetics.

## Course contents

Structure and taxonomy of viruses. Viral replication and genetics. Methods used for viral studies. Immune system response to virus. Effect of virus in cells. Vaccines and Antiviral Drugs. Laboratory Diagnosis of Viral Infection. Prions. Biology of animal cells. Equipment, consumables and reagents used in animal cell culture. Culture and subculture of animal cells: isolation, primary cultures and cell lines. Animal cell culture applications.

# Course contents (extended version)

- 1. General Characteristics, Structure and Taxonomy of Viruses
  - Viral Structure
    Viral Taxonomy
- Viral Taxonomy
   Cultivation and Characterization of Viruses
   Viral Propagation Methods
   Concentration and Purification of Viruses
   Infectivity and Storage
   Virus Visualisation.

  - Virus Visualization
     Direct Enumeration of Viruses

  - Indirect Enumeration of Viruses

    Miscellaneous Methods used for Characterization
- Viral Replication and Genetics
   Viral Replication of DNA and RNA viruses.
- Viral Replication of DNA and RNA viruses.
   Viral Genetics
   Interactions Between Two Viruses
   Virus Applications: Gene Therapy and Recombinant Vaccines
  4. Virus-Cell Interactions, Viral Pathogenesis and host defenses to viruses.
   Interaction Between Viruses and Host Cells
   Pathogenesis of Viral Infections
   Host Defenses
   Immunological Effects of Viral Infection.

  5. Prevention of Viral Diseases, Vaccines and Antiviral Drugs
   Vaccines
   Passive Immunization
- - Passive Immunization Herd Immunity
- Antiviral Drugs
   Laboratory Diagnosis of Viral Infections
   Diagnostic Approaches
   Virus Isolation

  - Virus Neutralization
     Protection Tests
- Protection Tests
   Collection and Submission of Specimens
   Families with Viruses of Veterinary Significance
   Circoviridae, Parvoviridae, Poxviridae, Herpesviridae, Papillomaviridae, Adenoviridae, Asfarviridae.
   Retroviridae, Reoviridae, Paramyxoviridae Rhabdoviridae, Orthomyxovirida, Picornaviridae
   Picornaviridae, Caliciviridae, Coronaviridae, Arteriviridae, Togaviridae and Flaviviridae.
   Bacteriophages, viroids and virusoids.
   Prions and Transmissible Spongiform Encephalopathies
   Prion Characteristics
   Transmissible Spongiform Encephalopathies of Animals

- - Transmissible Spongiform Encephalopathies of Animals
- Scrapie
   Bovine Spongiform Encephalopathy
   Feline Spongiform Encephalopathy
   Transmissible Encephalopathy of Mink
   Chronic Wasting Disease of Deer and Elk
   Spongiform Encephalopathy in Captive Ruminants
   Human Transmissible Spongiform Encephalopathies

  10. Culture of animal cells: basic technique.
   Animal cell biology
   Equipment consumples and reagents used in animal cells.

- Animal cell biology
   Equipment, consumables and reagents used in animal cell culture
   Culture and subculture of animal cells: isolation, primary cultures and cell lines
   Characterization and stipulation of cell lines
   Animal cell culture applications

# This document is valid only if stamped in all pages.

# Recommended reading

- Aiello, S., Moses, M., 2016. The Merck Veterinary Manual. 11<sup>a</sup> edição, Merck Elsevier Health Sciences.
   Barthold et al., 2011. Fenner's Veterinary Virology. 4<sup>a</sup> edição, Elsevier, Academic Press, EUA, 534 pp.
   Freshney, R., 2016. Culture of Animal Cells: A Manual of Basic Technique and Specialized Applications. 7<sup>a</sup> edição, John Wiley & Sons Inc, NY, EUA, 728 p.
   Johnson, A., 2014. Small Animal Pathology for Veterinary Technicians. John Wiley & Sons Inc NY, EUA, 240 pp.
   Quinn, P., Markey, B., Carter, M., Donnelly W., Leonard, F., 2011. Veterinary Microbiology and Microbial Diseases. Wiley-Blackwell, NJ, EUA, 928 pp.

# Teaching and learning methods

Lectures will be support by media and multimedia resources. Practical classes will engage direct working with animals and laboratory practices. Everyone is expected to contribute actively to discussions. Non present hours will involve training in a working environment. Graduate students are expected to work largely on their own initiative although with the close support and supervision of a tutor.

# Assessment methods

- 1. Coursework (Regular) (Final, Supplementary, Special)
   Intermediate Written Test 15% (1st written exam)
   Intermediate Written Test 15% (2nd written exam)
   Intermediate Written Test 30% (3th written exam)
   Reports and Guides 40% (1st written laboratory reports (10%)+2nd written laboratory reports(10%)+ 3th written reports (20%))
  2. final written exam (Student Worker) (Final, Supplementary, Special)
   Final Written Exam 60% (Final written exam)
   Final Written Exam 40% (written laboratory exam)

# Language of instruction

Portuguese

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06-11-2019	09-11-2019	10-11-2019	11-11-2019